

Decreased pericytes coverage in remodeled capillaries of chronically-rejected airway allografts. NG2 and α -SMA were used as pericyte markers. (**A**) Negative control stained with 2nd antibody only. (**B** and **C**) IF staining shows that remodeled, small caliber vessels in chronically-rejected day 56 allografts have more CD31⁺NG2⁻ (red; arrow) (**B**) and CD31⁺ α -SMA⁻ (green; arrow) vessels (**C**). Scale bar: 20 μ M.





Generation of lineage tracing model for Tie²⁺ cells. (**A**) Hemizygote mice expressing Tie²-Cre were crossed with homozygote mice expressing ROSA26EYFP to obtain (Tie²-Cre; ROSA26EYFP) double transgenic mice. (**B**) IF staining of the trachea from double transgenic mice show YFP⁺ cells express CD31, which is consistent with the majority of Tie²⁺ cells in a normal trachea being endothelial cells. Scale bar: 20μ M.







Recipient Tie²⁺ cells migrating into the rejecting allograft can become donor Pdgfrb⁺ pericytes. (**A**) IF staining of day 56 chronically-rejected airway allograft shows that YFP⁺ cell (green; arrow) expresses Pdgfrb(red; arrow). (**B**) Flow cytometry analysis of collagenase digested day 56 allografts confirms that YFP⁺ cells contain a population of Pdgfrb⁺ cells (left panel, no staining control; right panel, Pdgfrb staining). Scale bar: 20μ M.



Endothelial origin of Hif1 α and angiogenic growth factors, Sdf1, Vegf and Plgf. (**A**) IF staining of day 3 allografts (Balb/C to B6) shows that cells stained with nucleus-localized Hif1 α (green; arrow) express CD31. (**B-D**) IF staining of control day 0 trachea shows negative staining of Sdf1 (**B**), Vegf (**C**) and Plgf (**D**), staining of day 3 allografts shows that Sdf1 (**B**), Vegf (**C**) or Plgf (**D**) positive cells are stained with endothelial marker CD31 or Vegfr2. Scale bar: 20µM.



Generation of Hif1 α conditional knockout mice and characterization of HIF-1 α deficient airways. (**A**) Breeding strategy. (**B**) Mice carrying Cag-Cre-ER and Floxed Hif1 α genes were treated with tamoxifen for 5 consecutive days at a dose of 120mg/kg and waited for at least 2 more days to allow sufficient recombination. Genomic DNA was then extracted from the airways and subjected to PCR analysis. Tamoxifen treatment leads to disappearance of 950 bp product because of the excision of exon 2. Primers used for PCR: 5'-GCA GTT AAG AGC ACT AGT TG-3' and 5'-TGT TAA ATA AAA GCT TGG AC-3' as described in (1).



d8

Inhibition of CXCR4 mediated signaling by AMD3100 accelerated the loss of microvascular perfusion but did not affect the donor and recipient vascular anastomosis formation. AMD3100 treated allografts lost microvascular perfusion at day 8, and it did not affect the day 4 microvascular perfusion, which reflects the microvascular reconnection between the donor and the recipient. Scale Bar: 100µM.



d4 allografts

d8 allografts

d12 allografts

Supplemental Figure 7

LacZ expression kinetics of AdLacZ-treated airway allograft. AdLacZ-treated airway allografts were harvest at day 4 (**A**), day 8 (**B**) and day 12 (**C**). After X-gal staining (blue), the allografts were sectioned and subjected to H&E staining. Scale bar: 40μ M.

Supplemental Table 1. Sequences of primers for qRT-PCR analysis

| Gene | Forward Primer | Reverse Primer |
|--------|---------------------------|-------------------------------|
| Plgf | GGATGTGCTCTGTGAATGC | CCTCTGAGTGGCTGGTTAC |
| Vegf | GGCTGCTGTAACGATGAAG | CTCTCTATGTGCTGGCTTTG |
| Sdf1 | GAGAGCCACATCGCCAGAG | TTTCGGGTCAATGCACACTTG |
| Angpt1 | CTACCAACAACAACAGCATCC | CTCCCTTTAGCAAAACACCTTC |
| Angpt2 | CTGTGCGGAAATCTTCAAGTC | TGCCATCTTCTCGGTGTT |
| MCP-1 | GAAGGCCAGCCCAGCACCAG | GTGGATGCTCCAGCCGGCAA |
| IL-1β | GCCTCGTGCTGTCGGACCCA | TGAGGCCCAAGGCCACAGGT |
| IL-6 | GGTGACAACCACGGCCTTCCC | AAGCCTCCGACTTGTGAAGTGGT |
| Col1a1 | CCAGGCCCTGCCGGAGAAGA | CCAGGCCCTGCCGGAGAAGA |
| Col3a1 | CCAGAACATTACATACCACTGCAAA | GTGTTTAGTACAGCCATCCTCTAGAACTG |
| Col5a1 | GAGGCCACGAGACTAGAAGGAGGAG | CACGGCGCAGGGATCGACTGTG |
| 18S | GAATCGAACCCTGATTCCCCGTC | CGGCGACGACCCATTCGAAC |

Supplemental Data Reference:

1. Milosevic, J., Maisel, M., Wegner, F., Leuchtenberger, J., Wenger, R.H., Gerlach, M., Storch, A., and Schwarz, J. 2007. Lack of hypoxia-inducible factor-1 alpha impairs midbrain neural precursor cells involving vascular endothelial growth factor signaling. *J Neurosci* 27:412-421.