

Schematic diagram illustrating development of the vascular networks in the murine retina. The retinal vasculature forms as endothelial cells migrate from the optic nerve onto the retinal surface at birth and progress radially to form the superficial (or inner) plexus. Around postnatal day 7 (P7), sprouting vessels descend and advance into the OPL where they establish the deep plexus. At P11-12 stages, sprouting vessels from the deep plexus ascend into the IPL and ramify to form the intermediate plexus. GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer.



Putative neurovascular units in the INL. (**A**) Whole retinal sections from mice harboring two Cre-recombination reporters and *ptf1a-Cre*. (**B**) IHC for bipolar (Chx10; top panel) and Mueller glia (glutamine synthetase (GS); bottom panel) is shown to highlight that their locations in the INL are distinct from amacrine cells. (**C**) Cre-mediated recombination occurs in amacrine cells in the inner margin of the INL and colocalizes with a pan-amacrine cell marker, Syntaxin 1 (Stx1), a GABAergic amacrine cell marker (GAD), and glycinergic amacrine cell marker (Glyt1) in cryosectioned retinas. (**D**) The percentage of Cre-positive amacrine cells that colocalized with amacrine cell subtypes was determined by counting cells in cryosectioned retinas (≥250 cells were counted for each cell type). (**E**) Colocalization of Cre-mediated recombination (td-Tomato) with calbindin positive horizontal cells at P23. (**F** and **G**) IHC on thick cut sections on *ptf1a-Cre;R26<sup>GFP/+</sup>* mice with NF-M (**F**) and CD31 (**G**). (**H**) Immunofluorescence for anti-MAP-2 and anti-CD31 in cryosections from P23 *ptf1a-Cre*; R26<sup>tdTomato/+</sup> retinas. Scale bar: 50 µm (**A-G**); 20 µm (**H**). GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer.



The retinal vasculature of the superficial and deep plexuses are unaffected by *Vegfa* deletion in amacrine and horizontal cells. (**A**) Normal vascularization (bidirectional arrows) is observed in GS-lectin-positive P6 *ptf1a-Cre*; *VEGF*<sup>*i*/*t*</sup> retinas and controls (*VEGF*<sup>*i*/*t*</sup> (no Cre), *ptf1a-Cre*; *VEGF*<sup>*i*/*t*</sup> +). (**B-D**) Whole-mount staining in *VEGF*<sup>*i*/*t*</sup> or *ptf1a-Cre*; *VEGF*<sup>*i*/*t*</sup> retinas at P12 (**B**) or P15 (**C** and **D**), reveals no differences in the number of vertical sprouts descending through the IPL from the superficial plexus or ascending through the INL from the deep plexus (*n* = 4). Scale bar: 1 mm (**A**; upper panels); 100 µm (**A**; lower panels); 50 µm (**B** and **C**). (**E**) qPCR analysis showed that soluble VEGF120 and VEGF164 were the most abundant *Vegf* isoforms expressed in *ptf1a-Cre*; *VEGF*<sup>*i*/*t*</sup> mice at P15 (*n* = 4). SP; superficial plexus, IM; intermediate plexus, DP; deep plexus.



Chronic intermediate plexus attenuation is observed in ptf1a-Cre; *VEGF* knockout mice. The intermediate plexus capillaries in the whole mount retinas of *VEGF*<sup>//f</sup> or *ptf1a-Cre*; *VEGF*<sup>//f</sup> mice at 2, 4, 6, 12 months. Scale bar: 50 µm.

Figure-S4 (Friedlander)



Blood vessels do not advance to the OPL in *VhI* mutants (**A**) Vessels sprout from the superficial plexus towards the OPL in controls (*VHL*<sup>f/f</sup> mice), but are directed towards the IPL in *ptf1a-Cre*; *VHL*<sup>f/f</sup> mice at P5 where *Vegfa* is most highly expressed. (**B**) In vivo imaging of the ocular fundus and indocyanine green angiography in *ptf1a-Cre*; *VHL*<sup>f/f</sup> mice and controls revealed dense vasculature. (**C** and **D**) The dense convoluted intermediate plexus, and attenuated superficial and deep plexuses persisted until as late as 20 months (**C**). Note that the abnormally high number of branching points persists in both groups longitudinally (**D**) (n = 4-5). \*\*\**P*<0.001; 2-tailed Student's *t* tests. Error bars indicate mean ± SD. Scale bar: 50 µm (**A** and **C**). GCL, ganglion cell layer; INL, inner nuclear layer; ONL, outer nuclear layer.



Early angiogenesis events (P12) are regulated by VHL/HIF-1 $\alpha$ /VEGF signaling. (**A-F**) Combinatorial conditional knock-out strategies were employed to show that the loss of HIF-1 $\alpha$  (**B**) but not HIF-2 $\alpha$  (**C**) in amacrine and horizontal cells interferes with intermediate plexus development in haplosufficient Vhl<sup>+/-</sup> mutants compared with controls (**A**). (**D-F**) Homozygous deletion of Vhl and Hif-1 $\alpha$  (**E**) prevents the neovascularization observed in Vhl mutants, but deletion of HIF-2 $\alpha$  elicits no effect (**F**) compared with controls (**D**). (**G**) Homozygous deletion of Vhl and Vegfa also rescues the Vhl phenotype. (**H**) In situ hybridization for Vegfa in Vhl mutants and controls. (**J**) In situ hybridization for Vegfa in double Vhl/ Hif-1 $\alpha$  mutants and controls. (**J**) In situ hybridization for Vegfa in double Vhl/ Hif-2 $\alpha$  mutants and controls. (**K**) In situ hybridization for Vegfa in double Vhl/ Hif-2 $\alpha$  mutants and controls. (**K**) In situ hybridization for Vegfa in double Vhl/ Hif-2 $\alpha$  mutants and controls. (**K**) In situ hybridization for Vegfa in double Vhl/ Hif-2 $\alpha$  mutants and controls. (**K**) In situ hybridization for Vegfa in double Vhl/ Hif-2 $\alpha$  mutants and controls. (**L**) Relative mRNA expression values from qPCR gene-profiling analysis of 84 hypoxia signaling related genes in *ptf1a-Cre*; VHL<sup>t/f</sup>; VEGF<sup>t/f</sup> retinas at P12 compared with controls (harboring floxed alleles but no Cre); upregulated genes are plotted (n = 4). (**M**) Fold change of Vegfa isoforms in P12 *ptf1a-Cre*; VHL<sup>t/f</sup>; VEGF<sup>t/f</sup> retinas at P12 compared with controls (**A-K**).

Figure-S6 (Friedlander)



The genetic ablation of amacrine and horizontal cells in mice results in attenuation of the intermediate plexus and negatively affects visual function. (A) Schematic of the experimental design for the genetic depletion of amacrine and horizontal cells in ptf1a-Cre; R26<sup>DTR/+</sup> mice. (B) DT was injected daily from P6-8. (C and D) Thinning of the GCL/IPL and INL is observed in vivo using SD-OCT (C), and quantified (D) in P23 ptf1a-Cre; R26<sup>+/+</sup> and ptf1a-Cre; R26<sup>DTR/+</sup> mice (n = 6). (E) IHC from P23 DTtreated *ptf1a-Cre*; *R26*<sup>iDTR/+:tdTomato/+</sup> and *ptf1a-Cre*; *R26*<sup>+/+: tdTomato/+</sup> mice after DT treatment at P6-8 reveal a loss of the majority of amacrine and horizontal cells. (F) DT injections from P6-8 in P23 ptf1a-Cre; R26<sup>+/+</sup> and ptf1a-Cre; R26<sup>DTR/+</sup> staged mice induced attenuation of the intermediate plexus. (G) in situ hybridization was performed on P12 ptf1a-Cre; R26<sup>+/+</sup> or ptf1a-Cre; R26<sup>DTR/+</sup> retinas with a Vegfa probe and counterstained with DAPI. (H) Full-field ERGs performed on ptf1a-Cre; R26<sup>DTR/+</sup> mice 3 months after DT treatment at P30-32 revealed reduced b waves and gross defects in cone-driven pathways (n = 5-6). \*P<0.05. \*\*P<0.01. \*\*\*P<0.001; 2tailed Student's *t* tests. Error bars indicate mean  $\pm$  SD. Scale bar: 50  $\mu$ m (**F, G**); 40 μm (E). GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; ONL, outer nuclear layer.

Figure-S7 (Friedlander)



Visual acuity in ptf1a-Cre; VEGF knockout mice is significantly impaired. Optokinetic reflexes were measured in VEGF<sup>#</sup> or ptf1a-Cre; VEGF<sup>#</sup> mice and significant defects were observed in 3 month old mice (n = 10).

Figure-S8 (Friedlander)



Neurovascular units with the amacrine and horizontal cells and intraretinal capillaries in ptf1a-Cre; VEGF knockout and ptf1a-Cre; VHL knockout mice. Fluorescent immunostaining for anti neurofilament (NF) or anti-MAP2 with CD31 in retinal cryosections from P23 *ptf1a-Cre*; *VHL<sup>t/t</sup>* mice or *ptf1a-Cre*; *VEGF<sup>t/t</sup>* mice. The nuclei of cells were counterstained with DAPI (blue). Scale bar: 50 µm. GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; OPL, outer plexiform layer; ONL, outer nuclear layer.



No evidence of heightened neurodegeneration or abnormal synaptogenesis was observed in *ptf1a-Cre*; *VEGF*<sup>*i*/f</sup> mice. (**A**) TUNEL staining of P12 *ptf1a-Cre*; *VEGF*<sup>*i*/f</sup> retinas and controls. Scale bars: 500 µm (**A**; upper right); 50 µm (**A**; lower right). (**B**) Quantification from (**A**) at P12 or P23 (n = 4). (**C**) The expression pattern of synaptophysin is unremarkable in P15 *ptf1a-Cre*; *VEGF*<sup>*i*/f</sup> ;R26<sup>*t*/d</sup><sup>*Tomato/+*</sup> retinas. (**D**) Transmission electron micrograph showing normal synaptic ultrastructure (arrows) and synaptic vesicles in P23 *ptf1a-Cre*; *VEGF*<sup>*i*/f</sup> retinas (upper panels: controls). Scale bar: 50 µm (**C**); 1 µm (**D**). GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; ONL, outer nuclear layer; OPL, outer plexiform layer.

Figure-S10 (Friedlander)



No differences in the topographies of interneurons and Mueller glia are observed in ptf1a-Cre VEGF knockout and ptf1a-Cre; VHL knockout mice. (**A**-**L**) Cryosections from 23-day-old *ptf1a-Cre*; *VEGF<sup>#/#</sup>* and *ptf1a-Cre*; *VHL<sup>#/#</sup>* mice and controls were stained for calretinin (**A**-**C**), calbindin (**D**-**F**), chx10 (**G**-**I**), and glycogen synthase (GS) (**J**-**L**). The nuclei of cells were counterstained with DAPI (blue). Scale bar: 50 µm in **A**-**L** GCL, ganglion cell layer; IPL, inner plexiform layer; INL, inner nuclear layer; ONL, outer nuclear layer.

Figure-S11 (Friedlander)



Normal weight, blood glucose, and HbA<sub>1c</sub> levels in VEGF and VHL mutant mice. (**A**, **B**) Body weights in both groups were comparable. (n = 9-10). (**C**, **D**) Blood glucose was measured at P12 and P23 of *ptf1a-Cre*; *VEGF<sup>t/f</sup>* mice and *ptf1a-Cre*; *VHL*<sup>t/f</sup> mice. (n = 10 each). (**E**) HbA<sub>1C</sub> levels in 4 month-old *ptf1a-Cre*; *VEGF<sup>t/f</sup>* and 6 month-old *ptf1a-Cre*; *VHL*<sup>t/f</sup> mice. (n = 8 each).







Loss of VEGF in amacrine and horizontal cells accelerates photoreceptor atrophy in an animal model of retinal degeneration. (**A**) No degeneration (thinning of the ONL layer) is observed in SD-OCT images of P17 *ptf1a-Cre*; *VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/rd10</sup> mice compared with controls (*VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/rd10</sup>). (**B** and **C**) Significant ONL thinning is observed in rd10 mice with impaired intermediate plexuses (*ptf1a-Cre*; *VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/rd10</sup>) compared with rd10 mice (*VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/ rd10</sup>), or with non-degenerating controls (one recessive *rd10* allele; *ptf1a-Cre*; *VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/+</sup>) using OCT (**B**) and histology (**C**). (**D**) The integrity of the intermediate plexus is shown in P21 *ptf1a-Cre*; *VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/rd10</sup> mice (green) compared with controls (*VEGF<sup>1/f</sup>*; *Pde6b*<sup>rd10/rd10</sup>). Scale bar: 50 µm (**C**); 100 µm (**D-F**). IPL, inner plexiform layer; INL, inner nuclear layer; ONL, outer nuclear layer.

Figure-S13 (Friedlander)