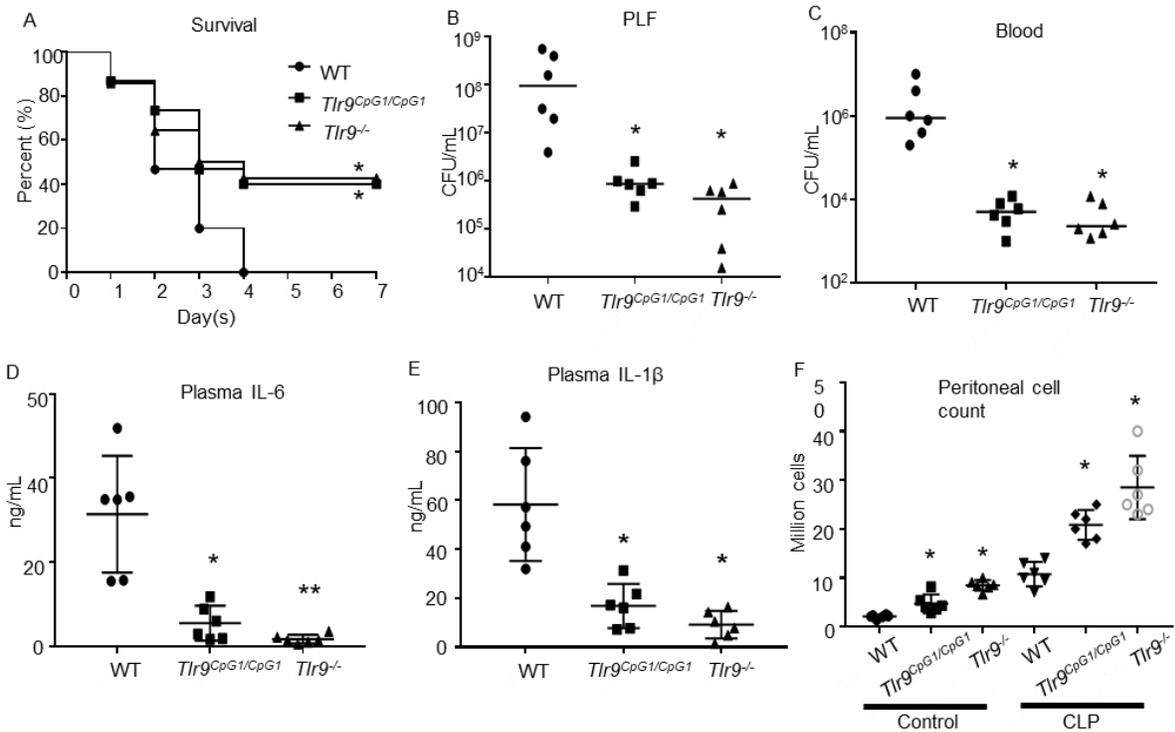
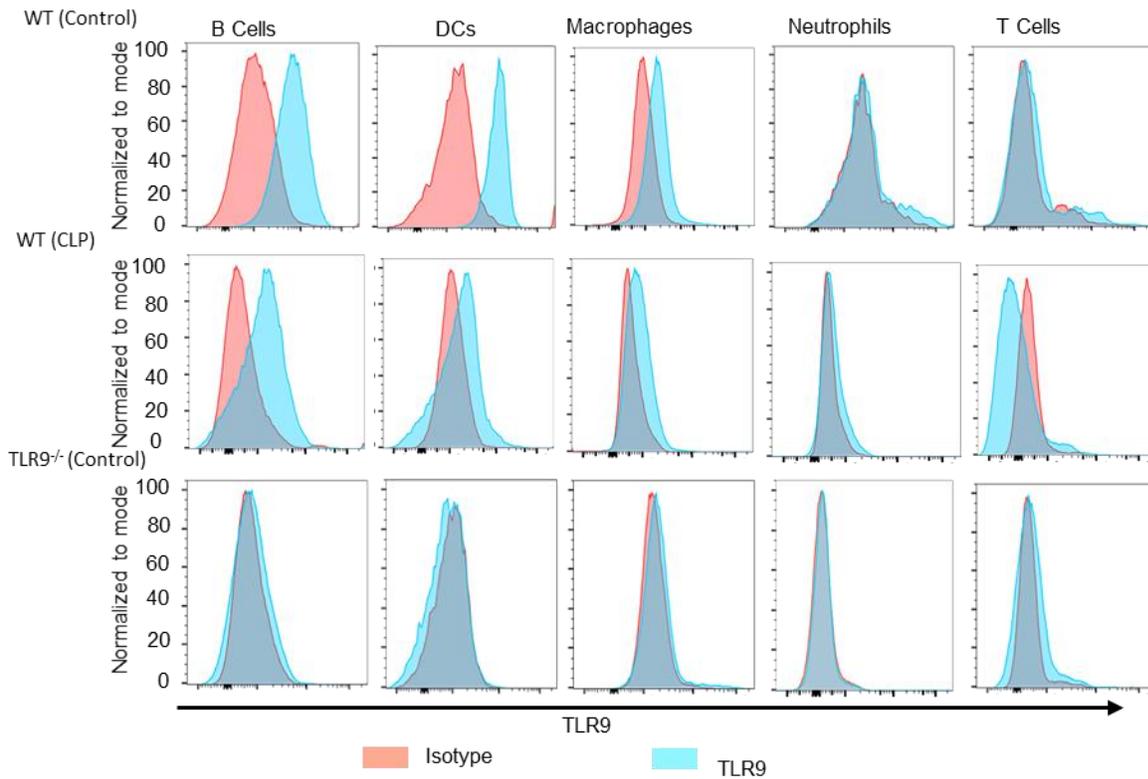


Supplemental Figure 1



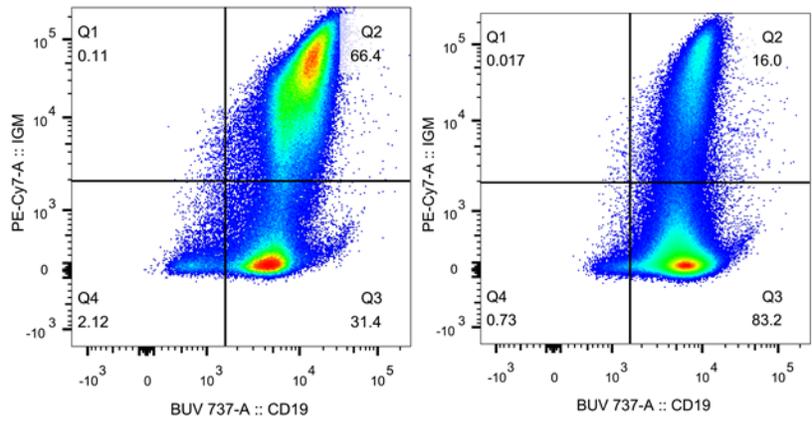
Supplemental Figure 1. Genetically blocking *Tlr9* signaling showed beneficial effects on sepsis. Wild-type (WT), *Tlr9^{CpG1/CpG1}*, and *Tlr9^{-/-}* mice were subjected to CLP. A: Seven-day survival after CLP. Data are from two separate experiments, n=20. Statistical difference was tested using the log-rank test. * $p < 0.05$. B-C: Blood and peritoneal lavage fluid (PLF) were collected at 18 hours after CLP. Bacterial counts in (B) peritoneal lavage fluid (PLF), and (C) blood. Data are from two separate experiments. Symbols represent for individual mice. Statistical difference was tested using a nonparametric Mann Whitney U statistic. * $p < 0.05$. D & E: Plasma cytokines levels. Blood was collected at 18 hours after CLP. Plasma (D) IL-6 and (E) IL-1 β concentrations were measured by ELISA. Data are means \pm SD from 2 separate experiments. Symbols represent for individual mice. Statistical difference was tested using student T test. * $p < 0.05$ vs WT. F: Peritoneal cell numbers were measured using cellometer. Data are means \pm SD from 2 separate experiments. Symbols represent for individual mice. Statistical difference was tested using 1- way ANOVA with Bonferroni's post hoc analysis, * $p < 0.05$, $p < 0.01$.

Supplemental Figure 2



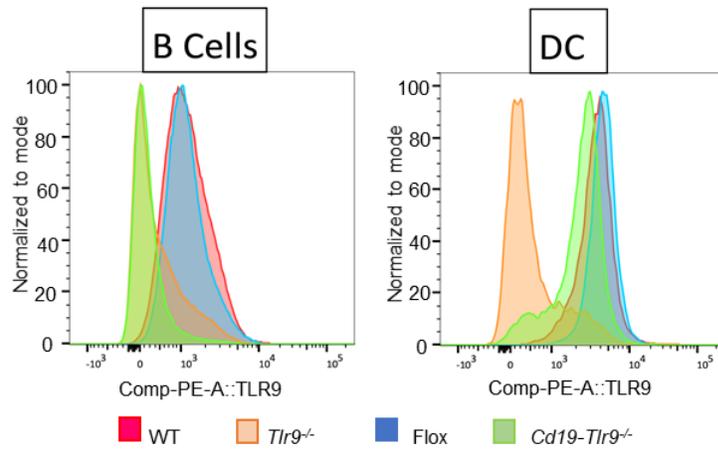
Supplemental Figure 2. TLR9 expression in peritoneal immune cell before and after CLP. WT and *Tlr9*^{-/-} mice were subjected to CLP. PLF was collected at 18 hours after CLP. TLR9 expression in indicated peritoneal cells. Mean Fluorescence Intensity (MFI) for TLR9 expression was measured by flow cytometry.

Supplemental Figure 3



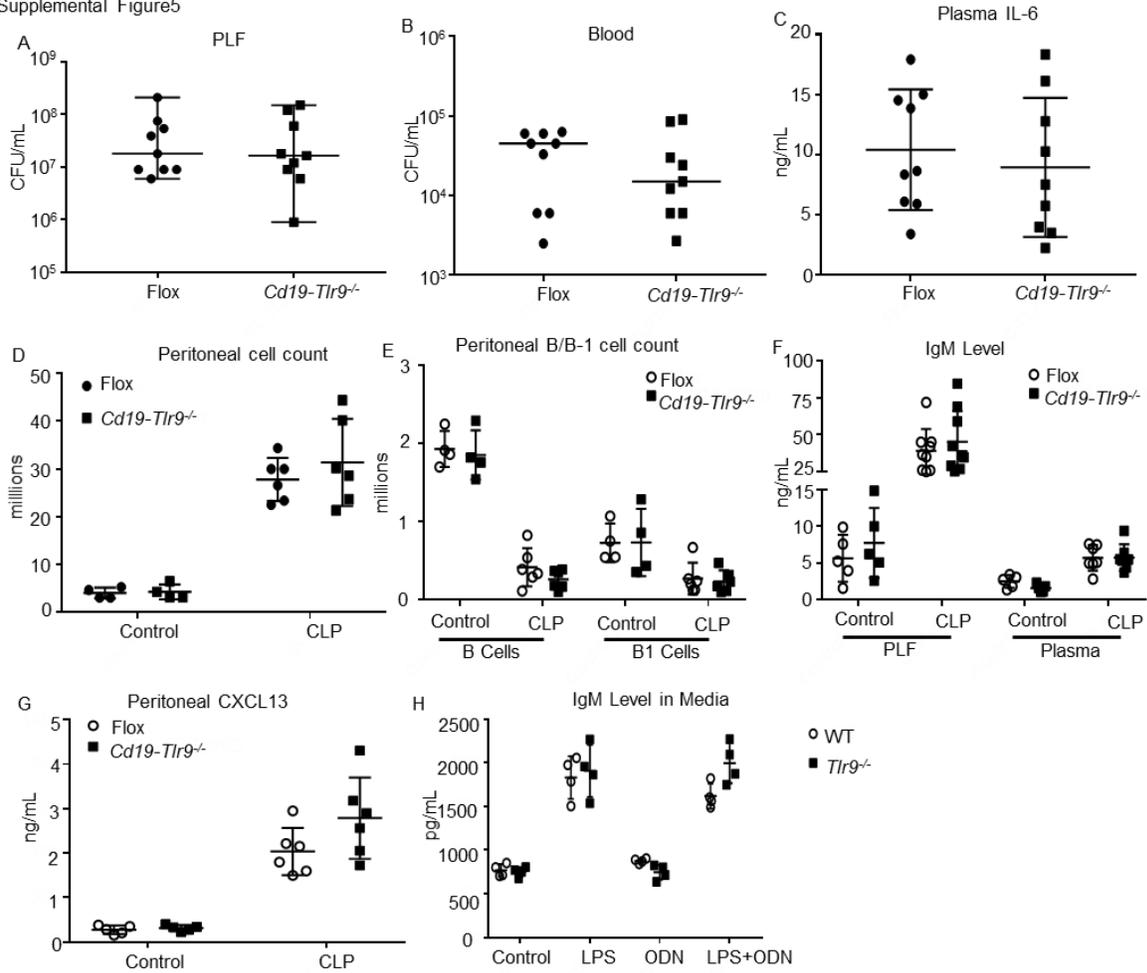
Supplemental Figure 3. Peritoneal B cells were depleted in *Tlr9*^{-/-} mice. *Tlr9*^{-/-} mice were treated with CD19 neutralizing antibodies (10mg/mouse) or control IgG for 24 hours. PLF was collected. Percent of peritoneal B cells was measured by flow cytometry

Supplemental Figure 4



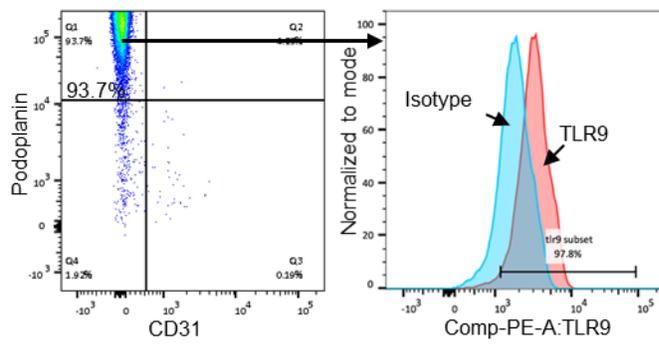
Supplemental Figure 4. Characterization of B cell specific *Tlr9*^{-/-} (*Cd19-Tlr9*^{-/-}) mice. WT, *Tlr9*^{-/-} mice were subjected to CLP without antibiotic treatment. PLF was collected from WT, *Tlr9*^{-/-}, Flox, and *Cd19-Tlr9*^{-/-} mice. TLR9 expression in B cells and DC was measured by flow cytometry.

Supplemental Figure 5



Supplemental figure 5. TLR9 in B cells is dispensable for sepsis. A-G: Flox and *Cd19-Tlr9^{-/-}* mice were subjected to CLP. Plasma and PLF was collected at 18 hours after CLP. Bacterial counts in (A) peritoneal lavage fluid (PLF), and (B) blood. Data are from two separate experiments. Symbols represent for individual mice. Statistical difference was tested using a nonparametric Mann Whitney U statistic. (C) Plasma IL-6 levels. (D) Peritoneal total cell count. (E) Peritoneal B cell and B-1 cell number; (F) Plasma IgM levels, (G) Peritoneal CXCL13 levels. Data are means \pm SD from 2 separate experiments. Symbols represent for individual mice. Statistical difference was tested using student T test. * $p < 0.05$. H: Sorted peritoneal B-1 cell were culture and treated with indicated TLR ligands (ODN1585: 5 μ M, LPS: 1 μ g/mL) for 18 hours. IgM levels in media was assessed using ELISA. Data are means \pm SD from 1 representative experiment. The experiments have been performed 3 times. Symbols represent for individual mice. Statistical difference was tested using unpaired, 2-tailed Student *t* tests.

Supplemental Figure 6



Supplemental Figure 6. TLR9 is constitutively expressed in mouse and human FRCs. The purity of cultured FRCs was assessed using flow cytometry. Numbers indicate percent of FRCs (CD45⁻, CD31⁻ and Podoplanin⁺). TLR9 expression in mouse FRCs.

Supplemental table1: Antibodies for flowcytometry

Species	Antibody	Company	Catalog Number
Mouse	Cd45-BUV395	BD	8037968
Mouse	Cd11b-PE	BD	557397
Mouse	F4/80-ef450	eBio	48-4801-82
Mouse	Ly6G-APC-CY7	BD	560600
Mouse	Cd11c-PE-CY5	eBio	15-0114-82
Mouse	MHCII-PE-CY7	invitrogen	4332615
Mouse	Cd19-BUV737	BD	564296
Mouse	Cd3e-FITC	eBio	11-0031-82
Mouse	Cd5-ef450	eBio	48-0051-82
Mouse	Cd34-PE	Biolegend	128609
Mouse	Tlr9-PE	BD	565640
Mouse	Tlr9 isotope-PE	BD	554880
Mouse	IgM-PE-CY7	eBio	25-5790-82
Mouse	Cd31PE-CY7	eBio	25-0311-81
Mouse	Podoplanin-APC	Biolegend	127410
Human	CD45-PE-CY5	BD	555484
Human	PODOPLANIN-BUV395	BD	747630
Human	CD31-APC-CY7	BD	563653
Human	TLR9-PE	BD	560425
Human	TLR9 isotope-PE	BD	554689

Supplemental table2: Primers for PCR

Species	Gene name	Forward	Reverse	Company
Mouse	Ccl2	GCATTAGCTTCAGATTACGGGT	TTAAAAACCTGGATCGGAACCAA	Invitrogen
Mouse	Ccl21	AAGGCAGTGATGGAGGGG	CGGGGTAAGAACAGGATTG	Invitrogen
Mouse	Ccl19	AGGTAGCGGAAGGCTTTCAC	CTGCTTCAGATTATCTGCCAT	Invitrogen
Mouse	Cxcl13	QT00107919		QIAGEN
Mouse	Cxcl2	CCAACCACCAGGCTACAGG	GCGTCACACTCAAGCTCTG	Invitrogen
Mouse	Cxcl3	QT00151599		QIAGEN
Mouse	Cxcl5	CGCTTCTTTCCACTGCGAGTGC	CTCAGTCATAGCCGCAACCGAGC	Invitrogen
Mouse	Gapdh	AACTTTGGCATTGTGGAAGG	ACACATTGGGGGTAGGAACA	Invitrogen
Human	CCL20	QT00012971		QIAGEN
Human	CXCL13	GCTTGAGGTGTAGATGTGTCC	CCCACGGGGCAAGATTTGAA	Invitrogen
Human	CXCL2	QT00013104		QIAGEN
Human	CXCL3	CGCCCAAACCGAAGTCATAG	GCTCCCTTGTTTCAGTATCTTTT	Invitrogen
Human	CXCL5	QT00203686		QIAGEN
Human	GAPDH	GAAGGTGAAGGTCGGAGTC	GAAGATGGTGATGGGATTC	Invitrogen